

# Comparison of antimicrobial effect of various oils mixed with zinc oxide – an *ex vivo, in vitro* study

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## ABSTRACT

**Background.** Success of endodontic treatment depends on complete elimination and various factors such as the antimicrobial activity of the obturating materials. Therefore a study was conducted to find the most prevalent species in the infected canals of primary teeth and to compare the antimicrobial effect of zinc oxide with various oils against most prevalent root canal pathogens.

**Materials and method.** The study was conducted on 100 children in the age group of 3 to 12 years with infected root canal in primary teeth. The sample was subjected to various microbiological analysis to identify the colonies. Following identification, the most common organism was taken and antimicrobial activity of zinc oxide eugenol, zinc oxide with coconut oil, zinc oxide with peppermint oil, zinc oxide with cinnamon oil, zinc oxide with tea tree oil on plates were tested. The zone of inhibition was measured and data were tested for statistical significance.

**Results.** *Enterococcus faecalis* was the most prevalent organism. It was seen that zinc oxide with tea tree oil had shown maximum antimicrobial activity against *Enterococcus faecalis* followed by zinc oxide with coconut oil, zinc oxide with peppermint oil, zinc oxide with cinnamon oil and zinc oxide with eugenol.

**Conclusions.** The study shows that *Enterococcus faecalis* was the most prevalent microbe in the infected root canals of primary molars. Zinc oxide with oil mixtures also has shown significant antimicrobial activity against *Enterococcus faecalis* when compared to zinc oxide eugenol.

**Keywords:** antimicrobial activity, zinc oxide, *Enterococcus faecalis*, *Streptococcus mutans*, essential oils

## INTRODUCTION

Oral cavity is a natural and favourable habitat for about 700 different types of organisms due to the presence of nutrients, epithelial debris and secretions. The imbalance due to oral environmental factors leads to changes in concentration of oral microbiomes and lead to diseases such as dental caries [1]. These bacteria invade through diseased tooth structures to the root and periapical tissue and has been implicated in the infections of endodontic origin [2].

The endodontic infection in deciduous teeth with pulp necrosis is of polymicrobial nature with predominance of anaerobic bacteria. However, these mi-

croorganisms disseminated in root canal system may persist even after cleaning and shaping of the root. The use of root canal filling materials with antibacterial properties should be able to eliminate residual pathogens, neutralize their toxic products and prevent canal reinfection in order to create a favourable environment for the healing process to proceed in periapical region [3,4]. It is therefore imperative that the endodontic filling materials used in primary teeth possess certain amount of antimicrobial activity.

Traditionally zinc oxide eugenol has long been used as an endodontic obturating material in primary tooth. Zinc oxide eugenol has been shown to cause slow resorption, irritation to oral tissues in children,

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### Article History:

Received: 5 May 2021  
Accepted: 13 May 2021

periapical tissue irritation, bone and cemental necrosis, affects the path of eruption of succedaneous tooth and has only limited antimicrobial action [5]. Therefore few researchers consider zinc oxide eugenol as an ideal root canal filling material [6]. So in search of a liquid that can replace eugenol, this study was undertaken to compare the antimicrobial action of zinc oxide with other oils such as coconut oil, cinnamon oil, peppermint oil, tea tree oil against the common root canal pathogen. Many natural oils like coconut oil, cinnamon oil, peppermint oil, tea tree oil have been used for treatment of various medical and dental problems since ancient times. They possess antibacterial, antifungal, and antioxidant properties [7,8].

Therefore this study was undertaken to find the predominant organisms in infected root canals and also to find the action of various formulations of zinc oxide with coconut oil, tea tree oil, peppermint oil and cinnamon oil against the most prevalent pathogenic organisms.

## MATERIALS AND METHODS

This *ex vivo-in vitro* study was done in the Department of Paediatric Dentistry of a Dental College in association with the Department of Microbiology of a Medical College. Before the start of the study institutional ethical committee approval (KDC/ETH/18/PED11/1) was taken.

The study consisted of two parts:

Step 1 – To identify most common organisms associated with infected primary root canal.

Step 2 – To assess the antimicrobial effect of zinc oxide with eugenol, zinc oxide with coconut oil, zinc oxide with peppermint oil, zinc oxide with cinnamon oil, and zinc oxide with tea tree oil against most common pathogens.

The study population comprised of 100 children aged between 3 to 12 years, having irreversible pulpal infections or infected root canals. The sample size was calculated using the formula  $n = Z_{1-\alpha/2} \cdot 2 \cdot SD \cdot d$   $Z_{1-\alpha/2} = 1.96$  at 95% confidence interval.  $SD$  = standard deviation of variable. Value of standard deviation can be taken from a previously done study.  $d$  = absolute error. Therefore the sample size  $= 1.96^2 (25)^2 = 96$ , rounded to 100 for study purpose. The inclusion criteria was children between 3 and 12 years with irreversible pulpal infection and infected root canal of primary lower molars. The exclusion criteria for the

study was children with oral candidiasis, children with any congenital or systemic or contagious diseases and children on antibiotics. The patient's parent /guardian was explained about the study and written informed consent was taken before the onset of the study.

After access opening, microbial samples were collected using a sterile paper point by passing it through the infected root canal of the primary molar for 30 seconds. Following removal from the canal, the paper point was immediately placed in cryovials and was transferred to microbiology department for analysis of various root canal pathogens. Lab procedures were conducted according to CLSI (Clinical and Laboratory Standard Institute) guidelines. The samples were inoculated on Blood Agar and Mac Conkey Agar plates and incubated at 37° C for 24 hrs. Laboratory procedures such as Gram staining and tests such as catalase test, salt tolerance test, oxidase test were done. Enterococcus group was confirmed through bile esculin test. Other colonies identified were confirmed through the Vitek test.

In the second part of the study, a standard inoculum was obtained by passing single colony of most prevalent microorganism in nutrient broth and the turbidity of broth was adjusted with McFarland 0.5 standard. By using lawn technique, microbial colonies were spread uniformly on Muller Hinton agar media. Then 6 wells each 4 mm in depth and 6mm in diameter were made in each of the agar plates with equal distance from each other. To each well zinc oxide with peppermint oil (Group 1), zinc oxide with tea tree oil (Group 2), zinc oxide with cinnamon oil (Group 3), zinc oxide with coconut oil (Group 4), zinc oxide with eugenol (Group 5) were placed. All the plates were kept in incubator at 37°C for 24 hrs and the diameter of zones of inhibition was measured and recorded.

## RESULTS

The specimens from 100 patients in the age group of 3 to 12 years were taken for this study. The patients constituted 62% females and 48% males.

Of the microorganisms isolated 55% were *Enterococcus faecalis* which is a Gram-positive facultative anaerobe, 30% were *Streptococcus mutans* which is a facultatively anaerobe, 5% were *Peptostreptococcus* which is an anaerobic, Gram-positive, non-spore forming bacteria, 6% were *Porphyromonas gingi-*

*valis* which is a black-pigmented, anaerobic, non-motile Gram-negative species, 4% were *Actinomyces* species which are gram-positive facultative anaerobes (Table 1).

**TABLE 1.** Prevalence of microorganisms from root canals of deciduous molar

| Species                         | Prevalence |
|---------------------------------|------------|
| <i>Enterococcus faecalis</i>    | 55%        |
| <i>Streptococcus mutans</i>     | 30%        |
| <i>Peptostreptococcus</i>       | 5%         |
| <i>Porphyromonas gingivalis</i> | 6%         |
| <i>Actinomyces</i>              | 4%         |

Figure 1 shows zone of inhibition on *Enterococcus faecalis*. Table 2 shows mean zone of inhibition values of different groups against *Enterococcus faecalis*. Diameters of zones of inhibition in mm for zinc oxide with tea tree oil paste were largest i.e.;  $41.1667 \pm 2.40$  followed by zinc oxide with coconut oil with a zone of inhibition of  $40.3333 \pm 1.50$  and zinc oxide with peppermint oil with a zone of inhibition of  $37.6667 \pm 2.06$  followed by zinc oxide with cinnamon oil with a zone of inhibition of  $36.5000 \pm 2.07$  and zinc oxide with eugenol paste was least with a zone of inhibition of  $29.0000 \pm 1.09$ .



**FIGURE 1.** Zone of inhibition of various oils mixed with zinc oxide powder against *Enterococcus faecalis*

**TABLE 2.** Mean zone of inhibition values of different groups against *Enterococcus faecalis*

| Groups  | Mean $\pm$ SD      |
|---------|--------------------|
| Group 1 | $37.6667 \pm 2.06$ |
| Group 2 | $41.1667 \pm 2.40$ |
| Group 3 | $36.5000 \pm 2.07$ |
| Group 4 | $40.3333 \pm 1.50$ |
| Group 5 | $29.0000 \pm 1.09$ |

Table 3 shows the antimicrobial effect using the One way ANOVA. The difference between and within the group was found to be highly statistically significant ( $p = 0.00$ )

**TABLE 3.** Antimicrobial effect of ZnO P, ZnO T, ZnO Ci, ZnO Co, ZnO E in the form of zone of inhibition on *Enterococcus faecalis* using One Way ANOVA

|                | Sum of Squares | df | Mean Square | F      | Sig. |
|----------------|----------------|----|-------------|--------|------|
| Between groups | 558.867        | 4  | 139.717     | 39.246 | .000 |
| Within groups  | 89.000         | 25 | 3.560       |        |      |
| Total          | 647.867        | 29 |             |        |      |

Table 4 shows the antimicrobial effect of zinc oxide with peppermint oil, zinc oxide with tea tree oil, zinc oxide with coconut oil, zinc oxide with cinnamon oil, zinc oxide with eugenol showing zone of inhibition on *Enterococcus faecalis* using Multiple Comparison Tukey's test. When antimicrobial effect of zinc oxide with peppermint oil was compared with zinc oxide with eugenol and zinc oxide with tea tree oil, it was found to be highly statistically significant ( $p < 0.05$ ). When zinc oxide with tea tree oil was compared with zinc oxide with cinnamon and zinc oxide with eugenol, the difference was found to be highly significant ( $p$  value = .000). The antimicrobial effect of zinc oxide with cinnamon was found to be statistically significant when compared with zinc oxide with coconut oil, zinc oxide with eugenol ( $p < 0.05$ ). Zinc oxide with coconut oil was found to be statistically significant when compared with zinc oxide with cinnamon and zinc oxide with eugenol ( $p = 0.05$ ). No statistical difference was found when zinc oxide with peppermint oil was compared with zinc oxide with cinnamon and zinc oxide with coconut oil.

## DISCUSSION

In the present study, teeth with infected root canal were selected. Microbial samples were collected using paper points placed in canals. Grossman et al. supported the use of sterile paper points for obtaining the root canal samples and showed that 1-10 microorganisms are optimally needed for obtaining growth through culture procedures [9].

The results of our study show the most common species isolated from infected root canals were *Enterococcus faecalis*. Colonies of *Streptococcus mutans*, *peptostreptococcus*, *porphyromonas gingivalis* and *actinomyces* species were also observed in the study

**TABLE 4.** Antimicrobial effect of ZnO P, ZnO T, ZnO Ci, ZnO Co, ZnO E in the form of zone of inhibition on *Enterococcus faecalis* using Multiple Comparison Tukey's Test

|           | (I)    | (J)    | Mean Difference (I-J) | Std. Error | Sig. | 95% Confidence Interval |             |
|-----------|--------|--------|-----------------------|------------|------|-------------------------|-------------|
|           |        |        |                       |            |      | Lower Bound             | Upper Bound |
| Tukey HSD | ZnO P  | ZnO T  | -3.50000*             | 1.08934    | .027 | -6.6993                 | -.3007      |
|           |        | ZnO Ci | 1.16667               | 1.08934    | .819 | -2.0326                 | 4.3659      |
|           |        | ZnO Co | -2.66667              | 1.08934    | .136 | -5.8659                 | .5326       |
|           |        | ZnO E  | 8.66667*              | 1.08934    | .000 | 5.4674                  | 11.8659     |
|           | ZnO T  | ZnO P  | 3.50000*              | 1.08934    | .027 | .3007                   | 6.6993      |
|           |        | ZnO Ci | 4.66667*              | 1.08934    | .002 | 1.4674                  | 7.8659      |
|           |        | ZnO Co | .83333                | 1.08934    | .938 | -2.3659                 | 4.0326      |
|           |        | ZnO E  | 12.16667*             | 1.08934    | .000 | 8.9674                  | 15.3659     |
|           | ZnO Ci | ZnO P  | -1.16667              | 1.08934    | .819 | -4.3659                 | 2.0326      |
|           |        | ZnO T  | -4.66667*             | 1.08934    | .002 | -7.8659                 | -1.4674     |
|           |        | ZnO Co | -3.83333*             | 1.08934    | .013 | -7.0326                 | -.6341      |
|           |        | ZnO E  | 7.50000*              | 1.08934    | .000 | 4.3007                  | 10.6993     |
|           | ZnO Co | ZnO P  | 2.66667               | 1.08934    | .136 | -.5326                  | 5.8659      |
|           |        | ZnO T  | -.83333               | 1.08934    | .938 | -4.0326                 | 2.3659      |
|           |        | ZnO Ci | 3.83333*              | 1.08934    | .013 | .6341                   | 7.0326      |
|           |        | ZnO E  | 11.33333*             | 1.08934    | .000 | 8.1341                  | 14.5326     |
|           | ZnO E  | ZnO P  | -8.66667*             | 1.08934    | .000 | -11.8659                | -5.4674     |
|           |        | ZnO T  | -12.16667*            | 1.08934    | .000 | -15.3659                | -8.9674     |
|           |        | ZnO Ci | -7.50000*             | 1.08934    | .000 | -10.6993                | -4.3007     |
|           |        | ZnO Co | -11.33333*            | 1.08934    | .000 | -14.5326                | -8.1341     |

which confirms the polymicrobial nature of colonization of infected root canals of deciduous molars. The microorganisms of the root canal have several characteristics leading to biologic and pathogenic events such as antigenicity, mitogenic characteristics, chemotaxis, histolysis with enzymes and activation of host cells [10].

Various studies have analysed the composition of microorganisms in the root canal with a persistent apical lesion. The analysis of these studies showed varied predominance of different species [11]. Pinheiro et al. found in their study that, *Enterococcus faecalis* was the most frequently recovered bacterial species among the facultative anaerobic species and gram positive microorganisms [12].

*Enterococcus faecalis* is occasionally found in cases of primary endodontic infections as well as in failed endodontic infections [13]. Enterococci survive in root canal infections where there are less nutrients and are resistant to the antimicrobial agents due to an effective proton pump mechanism which maintains optimal cytoplasmic pH levels [11]. Enterococci has the natural ability to acquire, gather and has ability for encoding virulence traits of extra chromosomal elements paving way to colonize, it also competes with other bacteria, and resist host defence

mechanisms leading to pathological changes. These changes may be initiated on production of toxins or indirectly through induction of inflammation [14,15]. The pioneering studies by Sundqvist [16] and later by Love et al. [17] demonstrated that, in addition to enterococci, *streptococci*, *lactobacilli*, and *Actinomyces*, obligately anaerobic species of *Fusobacterium*, *Eubacterium* spp., *Propionibacterium* spp., *Bifidobacterium* spp., *Peptostreptococcus micros*, and *Veillonella* species dominated the root canal microflora. Facultative anaerobes belong to the viridans group Streptococci and are commonly implicated in dental abscess. The viridans group Streptococci includes the mutans group [18].

Brook et al. found that the presence of *Prevotella*, *Porphyromonas*, *Fusobacterium* and *Peptostreptococcus* in pulpitis and dentoalveolar abscess [19]. This is in accordance to our study in which few colonies of *Porphyromonas* and *Peptostreptococcus* were seen.

It is widely known that the plant oils possess certain medicinal properties and studies have been actively conducted to prove its antimicrobial efficiency. Therefore in the present study antimicrobial effect of zinc oxide with tea tree oil, zinc oxide with cinnamon oil, zinc oxide with peppermint oil and zinc oxide with coconut oil and zinc oxide with eugenol was

evaluated against *Enterococcus faecalis* which was the predominant organisms detected in our study.

The components of plant oils are known to interfere with the enzymatic activity, thereby destroying bacterial leading to antimicrobial action. They also block clusters of Gram-positive bacteria, slow down bacterial replication, and remove endotoxin from Gram-negative bacteria to reduce oral pathogenic diseases [20].

In the present study, Agar diffusion method was used. It is one of the most commonly employed technique for evaluation of antimicrobial activity [21]. The results show that zinc oxide with tree tea oil has strong inhibitory properties as compared to routinely used zinc oxide eugenol. Our results which are in agreement to the findings of the study by Thosar et al. in 2018 [22] which shows that the tree tea oil with zinc oxide has a better antimicrobial action when compared with zinc oxide eugenol. Another study done by Thosar et al. [23] on five essential oils showed that tree tea oil has significant inhibitory effect on *Enterococcus faecalis*.

Various studies show the antimicrobial effect of tree tea oil. Ghayathri et al. [24] and Stoica et al. [25] have also shown the significant inhibitory effect of tea tree oil on *Enterococcus faecalis*. The antibacterial effect of tea tree oil is due to terpinen-4-ol,  $\alpha$ -terpinol and 1,8-cineole. Terpinen-4-ol enters the cell membrane of microorganisms and acts against its structural permeability. This in turn affects the metabolism of certain microorganisms leading to bactericidal and fungicidal effects [7].

Our study showed significant zone of inhibition by zinc oxide with coconut oil against *Enterococcus faecalis*. There are no studies in literature about the antimicrobial effect of zinc oxide with coconut oil. The antibacterial property of coconut oil is because of the presence of medium chain fatty acids [26]. Devan et al. [27] have shown in 2019 that medium chain fatty acids when used as a root canal irrigant has a significant action against *Enterococcus faecalis*. Studies show that lauric acid which is constituent of coconut oil exhibits significant antimicrobial activity against *E. faecalis* [28]. The inhibitory action of coconut oil may be due to their surfactant activity and their ability to cause cellular lysis by disrupting cell membranes by specific interaction with sites

within the microorganism or nonspecific interaction inhibiting physiological function [29].

Zinc oxide with pepper mint oil also showed significant antibacterial action. Thosar et al. [23] in a study against *Enterococcus faecalis* found that peppermint can act as an effective intracanal antiseptic solution against oral pathogens. Manoj et al. found that peppermint oil has significant action against *Enterococcus faecalis* [31]. Peppermint oil is obtained from the stem, leaves, and flowers of *Mentha piperita* plant by steam distillation method. Its principal constituents include monoterpinic alcohols mainly menthol, ketones mainly menthones, some monoterpenes and oxides. Other active constituents are menthol, menthone, cineol, and several other volatile oils [32].

In the present study zinc oxide with cinnamon oil has shown to have antimicrobial property against *Enterococcus faecalis*. Panchal et al. [33] compared the antibacterial efficacy of cinnamon extract and calcium hydroxide as an intracanal medicament against *Enterococcus faecalis* and found that cinnamon extract showed good antimicrobial efficacy. The antibacterial activity of cinnamon leaf extract against *Enterococcus faecalis* is shown to have inhibition zones at 20% concentration. Cinnamaldehyde, the most abundant component of cinnamon oil is a phenylpropanoid that has proven activity against microorganisms [34,35].

### Limitations of the study

Sample size for the study was fewer for species identification. The use of advanced technique such as polymerised chain reaction, DNA-DNA hybridisation etc would have given more specificity and sensitivity to the species identification. Many other factors such as saliva, mucus layer, clearance capacity, blood flow and normal flora could have had an effect on the results of this study. The antimicrobial effect of the materials was done in vitro in laboratory conditions where intra oral conditions are dynamic with lot of factors involved in antimicrobial action. Thus there is a need for more *in vivo* studies to standardize the microbial activity. Further research is also required to elucidate the mechanistic details as well as to evaluate the toxicity and clinical efficacy.

## CONCLUSIONS

In this study we found that gram positive facultative anaerobes are the most predominant species. *Enterococcus faecalis* was found to be the most predominant followed by *Streptococcus mutans*, *peptostreptococcus*, *porphyromonas gingivalis* and *actinomyces species*. All zinc oxide with oil mixtures

has shown antimicrobial activity against *Enterococcus faecalis* predominance. Hence, this study concludes that apart from traditional use of eugenol, oils such as peppermint oil, tea tree oil, coconut oil and cinnamon oil with zinc oxide can be considered as root canal obturating materials in primary teeth due to their antimicrobial properties.

*Conflict of interest:* none declared

*Financial support:* none declared

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